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## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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<b>(21) International Application Number:</b> PCT/EP92/01261 <b>(22) International Filing Date:</b> 5 June 1992 (05.06.92) <b>(30) Priority data:</b> MI91A001644 14 June 1991 (14.06.91) IT <b>(71) Applicant (for all designated States except US):</b> ITALFARM-ACO S.P.A. [IT/IT]; Viale Fulvio Testi, 330, I-20126 Milano (IT). <b>(72) Inventors; and</b> <b>(75) Inventors/Applicants (for US only) :</b> CARAMAZZA, Ida [IT/IT]; ZOPPETTI, Giorgio [IT/IT]; MODENA, Daniela [IT/IT]; SICCARDI, Antonio, G. [IT/IT]; BERETTA, Alberto [IT/IT]; LOPALCO, Lucia [IT/IT]; Via G. Carducci, 125, I-20099 Sesto San Giovanni (IT).		<b>(74) Agent:</b> MINOJA, Fabrizio; Studio Consulenza Brevettuale, Via Rossini, 8, I-20122 Milano (IT). <b>(81) Designated States:</b> AT (European patent), AU, BB, BE (European patent), BF (OAPI patent), BG, BJ (OAPI patent), BR, CA, CF (OAPI patent), CG (OAPI patent), CH (European patent), CI (OAPI patent), CM (OAPI patent), CS, DE (European patent), DK (European patent), ES (European patent), FI, FR (European patent), GA (OAPI patent), GB (European patent), GN (OAPI patent), GR (European patent), HU, IT (European patent), JP, KP, KR, LK, LU (European patent), MC (European patent), MG, ML (OAPI patent), MN, MR (OAPI patent), MW, NL (European patent), NO, PL, RO, RU, SD, SE (European patent), SN (OAPI patent), TD (OAPI patent), TG (OAPI patent), US.  <b>Published</b> <i>With international search report.</i>
<b>(54) Title:</b> ACYLATED HEPARINS AS INHIBITORS OF THE REPLICATION OF RETROVIRUSES  <b>(57) Abstract</b> <p>Heparins that have been N-acylated with mono- or dicarboxylic acids inhibit the replication of retroviruses, in particular HIV.</p>		

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WO 92/22588

PCT/EP92/01261

ACYLATED HEPARINS AS INHIBITORS OF THE REPLICATION OF  
RETROVIRUSES

The subject of the present invention is N-acylated heparins that are capable of inhibiting the replication of retroviruses, in particular that of the human immunodeficiency virus (HIV).

5        These N-acylated heparins have a level of acylation, defined as the percentage ratio of the number of N-acylated residues to the number of N-sulphate groups present in the original heparin, of between approximately 10% and approximately 100%. They  
10       are prepared from heparins with molecular weights ranging between approximately 1,000 and approximately 30,000 Daltons, in other words commercial heparins of extractive origin, or fractions or fragments of such heparins.

15       The subject of the present invention is therefore heparins that have been N-acylated with residues derived from aliphatic monocarboxylic acids with between 3 and 20 carbon atoms, such as propionic, butyric, caproic, caprylic, caprinic, lauric, myristic,  
20       palmitic, stearic, crotonic, oleic, elaidinic, stearolic, tetrolic, arachic and similar acids, or with residues of aliphatic dicarboxylic acids containing between 3 and 10 carbon atoms, such as malonic, succinic, dimethylmalonic, glutaric, suberic, azelaic  
25       and sebacic acids. These mono-or di-carboxylic acids can also be substituted by one or more hydroxyl, amine or acylamine groups, to give, for example, heparins that are N-acylated with residues of lactic,

WO 92/22588

PCT/EP92/01261

2

hydracrylic, tartaric, tartronic, malic, aspartic, glutamic, N-acetylaspartic, N-acetylglutamic and similar acids.

5 N-succinylated heparins containing between 25 and 35% of succinic residues are described in US Patent No. 3,118,816, and heparins with a level of acylation of approximately 100% are known from GB Patent No. 2,078,768.

10 Hence a further subject of the present invention is the use of these heparins as agents capable of inhibiting the replication of retroviruses, in particular that of the human immunodeficiency virus (HIV).

15 The inhibition of HIV replication by sulphated polysaccharides has been the subject of in-depth studies for some time (Lancet, 1, 1379, 1987; Biochem. Pharmacol. 37, 2987, 1988; Proc. Natl. Acad. Sci. U.S.A. 85, 6132, 1988). The polysaccharides that have been considered so far mainly belong to two classes: on 20 the one hand, polysaccharides of bacterial or vegetable origin that are sulphated either naturally or artificially, such as carrageenans, pentosan sulphate, dextran sulphate, xylan sulphate (EP-A-0,406,685), and on the other hand, heparins or heparin fragments or 25 fractions (EP-A-0,355,905). The former are believed to have the advantage of having little or no anticoagulant activity but little is known about their possible toxic effects, whereas the latter interfere significantly with the processes of blood coagulation but have the 30 advantage of not carrying any unknown risks as regards their therapeutic use since they have already been used

WO 92/22588

PCT/EP92/01261

3

in medical practice for some considerable time.

The therapeutic value of non-heparin sulphated polysaccharides was recently explained on the basis of their ability to prevent the glycoprotein gp120 expressed on the surface of infected cells from interacting with the CD4 receptor of healthy cells, thereby inhibiting the subsequent formation of specific cell structures (syncytia) that play a primary role in the spread of the infection (J. Acquired Immun. Deficiency Syndromes, 3, 493, 1990).

It has now been found that the N-acylated heparins described above are highly effective in inhibiting the replication of retroviruses, in particular that of HIV, produce few, if any, signs of toxicity and have virtually no anticoagulant or antithrombotic activity.

The N-acylated heparins that are the subject of the present invention are obtained by a process that comprises N-desulphation of the initial heparins by known methods and their subsequent acylation by means of reaction with the chosen acid or one of its functional derivatives.

The N-desulphation is achieved in particular by heating solutions of the chosen heparin in the form of the corresponding heparinic acid to temperatures of between 30 and 60°C for approximately 24 hours. The level of N-desulphation and therefore of N-acylation that can be obtained depends on the reaction temperature used. Thus, for example, a temperature of approximately 55-60°C for 24 hours produces a level of N-desulphation of over 95%, whereas a temperature of not more than approximately 35°C gives a level of N-

WO 92/22588

PCT/EP92/01261

4

desulphation of approximately 30%. Temperatures between these two values obviously give N-desulphation values between the two limits given above. The subsequent N-acylation reaction is carried out using preferably, as the acylating agent, a symmetrical or mixed anhydride of the chosen acid. The reaction is effected for the most part at ambient temperature, with the pH of the solution being kept at between approximately 5 and approximately 7 by the addition of a suitable alkaline agent. The final product is thus obtained in the form of a salt, preferably the salt of an alkali or alkaline earth metal, which can be further purified or converted into another salt, preferably the salt of an alkali or alkaline earth metal, by methods known to experts in the subject.

By way of example, we describe below the preparation of a number of heparins with varying levels of N-succinylation, obtained by starting with heparins of different molecular weights.

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#### EXAMPLE 1

##### Synthesis of N-succinylated heparin (100%)

###### a) Preparation of N-desulphated heparin.

The N-desulphated heparin was prepared using a modified version of the method already described (Lloyd et al., Biochem. Pharm., vol. 30, 637-648, 1971). An aqueous solution containing 1.1 g porcine sodium heparin with a mean molecular weight of 13,600 D, with anti-Xa activity of 206 units/mg (chromogenic) and APTT activity of 202 units/mg, is percolated through a column thermostatically maintained at 4°C containing 50 ml Dowex<sup>®</sup> 50 W X 8H cationic resin. The eluate (50 ml)

WO 92/22588

PCT/EP92/01261

5

containing the heparinic acid has a pH of approximately 1.2. The solution thus obtained is placed in a flask and kept at 55°C for 24 hours, with gentle stirring and continuous monitoring of the temperature. The solution is then neutralised to pH 7 with 1 N NaOH at ambient temperature. The solution is then freeze-dried, giving the N-desulphated heparin in the form of a white powder. The level of N-desulphation is >95% according to  $^{13}\text{C}$ -NMR analysis (shifting of the  $\text{C}_1$  signal of the aminosugar from 99.5 to 94 ppm and shifting of the  $\text{C}_2$  signal from 60.5 to 66.8 ppm).

b) Preparation of N-succinylated heparin.

The intermediate product obtained in a) above is dissolved in 100 ml water (1% solution) with stirring. The solution is neutralised to pH 7 with 0.1 N NaOH, then 2 g solid succinic anhydride are added in aliquots of 250 mg at a time, the entire procedure being carried out at ambient temperature. After each addition it is necessary to readjust the pH from 5 to 7 with 0.1N NaOH. At the end of the reaction the excess insoluble sodium succinate is filtered off. The clear solution thus obtained is treated with 3 volumes of absolute ethanol, to produce a precipitate. The precipitate is washed repeatedly with absolute ethanol. A solid, white precipitate is obtained. The precipitate is redissolved in water (5% solution). The solution is placed in a dialysis membrane (SPECTRAPOR<sup>®</sup> cut-off 8000 D) for 24 hours against 2 l of water. At the end of this time, the intradialysis solution is freeze-dried to give the N-succinylated heparin in the form of a white powder, which has the characteristics set out in Table 1.

WO 92/22588

PCT/EP92/01261

6

EXAMPLE 2Synthesis of partially (50%) N-succinylated heparin

The procedure is the same as that described in Example 1 but uses for the succinylation the partially  
5 (>50%) N-desulphated heparin derivative which can be obtained by the method described in Example 1a, though in this case the reaction is carried out at 40°C for 24 hours. The level of desulphation of this product is checked by the <sup>13</sup>C-NMR test as already described. The  
10 partially (50%) N-succinylated heparin in the form of a white powder has the characteristics set out in Table 1.

EXAMPLE 3Synthesis of partially (30%) succinylated heparin

15 The procedure is the same as that described in Example 1 but uses for the succinylation the partially (>30%) N-desulphated heparin derivative which can be obtained by the method described in Example 1a, though in this case the reaction is carried out at 35°C for 24  
20 hours. The partially (30%) N-succinylated heparin in the form of a white powder has the characteristics set out in Table 1.

EXAMPLE 4Synthesis of 100% N-succinylated LMW heparin

25 The procedure is the same as that described in Example 1, but uses 1 g of low molecular weight heparin (average molecular weight 5000 D), with anti-Xa activity of 97 units/mg and APTT activity of 74 units/mg (Calbiochem). In this case the N-succinylated  
30 derivative is purified from the excess succinate in a dialysis membrane (Spectrapor®) with a cut-off of 1000

WO 92/22588

PCT/EP92/01261

7

D, again as already described in Example 1. After freeze-drying, the 100% N-succinylated LMW heparin is obtained in the form of a white powder, which has the characteristics set out in Table 1.

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#### EXAMPLE 5

##### Synthesis of partially (50%) N-succinylated LMW heparin

The procedure is the same as that described in Example 2, and uses 1 g of the same low molecular weight heparin as used in the previous example. This yields 50% of N-succinylated LMW heparin with the characteristics set out in Table 1.

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#### EXAMPLE 6

##### Synthesis of 100% N-succinylated VLMW heparin

The procedure is the same as that described in Example 1, but uses 1 g of very low molecular weight heparin (molecular weight <3000 D, anti-Xa activity: >50 units/mg; APTT activity: <45 units/mg) (Sigma). In this case the N-succinylated derivative is purified from the excess succinate by means of fractional precipitation, which is based on the selective formation of complexes with alkylammonium salts using the original method of Scott (Methods of Biochem. Anal., vol. 8, 145-197, 1960). In brief, the 5% solution of N-succinylated VLMW heparin is subjected to selective precipitation using 10% cetylpyridinium chloride in the absence of salts, in the cold (4°C); the mixture is left to stand for approximately 2 hours, then centrifuged at 4000 rpm for 20 minutes. The resulting precipitate is separated from the excess succinate that remains in solution. After being washed 3 times with hot, distilled water, the precipitate is

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WO 92/22588

PCT/EP92/01261

8

redissolved hot (30-35°) in a 2M NaCl solution. Finally, the solution is treated with 3 volumes of absolute ethanol, causing the precipitation of the N-succinylated VLMW sodium heparin, which has the characteristics set out in Table 1.

#### EXAMPLE 7

##### Synthesis of partially (50%) N-succinylated VLMW heparin

The procedure is the same as that described in Example 2 and uses 1 g of the same very low molecular weight heparin used in the previous example. The N-succinylated derivative is purified from the excess succinate in the same way as described in Example 6. This yields 50% N-succinylated VLMW heparin, which has the characteristics and biological activity set out in Table 1.

#### EXAMPLE 8

##### Calcium salt of partially (50%) N-succinylated heparin

The product of Example 2 (1 g) is percolated through a column thermostatically maintained at 4°C and containing 50 ml DOWEX 50W-X8H<sup>+</sup> cationic resin. The eluate containing the acid form of the intermediate product is titrated with a stoichiometric quantity of calcium carbonate (CaCO<sub>3</sub>). The solution is then freeze-dried, yielding the partially (50%) N-succinylated calcium heparin in the form of a white powder, which has the characteristics set out in Table 1.

#### EXAMPLE 9

##### Calcium salt of partially (50%) N-succinylated LMW heparin

The product of Example 5 is treated as described

WO 92/22588

PCT/EP92/01261

9

in Example 8 to obtain the calcium salt of the partially (50%) N-succinylated LMW heparin, which has the characteristics set out in Table 1.

EXAMPLE 10

5 Calcium salt of partially (50%) N-succinylated VLMW heparin

The product of Example 7 is treated as described in Example 8 to obtain the calcium salt of the partially (50%) N-succinylated VLMW heparin, which has  
10 the characteristics set out in Table 1.

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WO 92/22588

PCT/EP92/01261

10

TABLE 1

Heparin derivatives	Carboxyl (1) sulphates	EF (2) (Baacetate) $R_F$	EF (3) (HCl) $R_F$	Ava (4) U/mg	APTT (5) U/mg	MW (6) (HPLC)
Sodium heparin	2.14	2 stains	$R_F=0.72$	206.0	202.0	13300
Ex. 1	0.91	single stain	$R_F=0.61$	< 1	26.4	13300
Ex. 2	1.2	2 stains	$R_F=0.64$	< 1	18.3	13030
Ex. 3	1.44	2 stains	$R_F=0.68$	19.5	74.8	13300
Ex. 4	1	single stain	$R_F=0.62$	< 1	4.9	4070
Ex. 5	1.1	single stain	$R_F=0.66$	< 1	< 1	3660
Ex. 6	0.9	single stain	$R_F=0.74$	< 1	2.4	2350
Ex. 7	1.1	single stain	$R_F=0.78$	< 1	< 1	2400
Ex. 8	1.2	2 stains	$R_F=0.64$	< 1	19	13300
Ex. 9	1.1	single stain	$R_F=0.74$	< 1	< 1	3660
Ex. 10	1.1	single stain	$R_F=0.78$	< 1	< 1	2400

1. Carb. Res. 39,168,1978
2. J. Chron. 196,398,1983
3. Anal. Biochem. 41,67,1971
4. Thromb. Res. 8,413,1976
5. Thromb. Res. 39,640,1978
6. J. Chron. 261,287,1983

WO 92/22588

PCT/EP92/01261

11

The antiviral activity of the compounds described above was evaluated with the aid of the syncytia formation inhibition test, using the method described below.

5           8E51 cells permanently infected with LAV+ (Lymphotropic Adenopathy Virus) and defective in the pol gene, and MOLT3 cells (CD4+ human T-lymphoblastoid line) cultivated in RPMI 1640 medium with 10% foetal calf serum in a concentration of  $10^6$  cells/ml are  
10 incubated together in a ratio of 1:2 and distributed in Microtiter plates with 96 conical-bottomed wells. The heparin derivatives are added to the cells in the appropriate dilutions, then after 30 minutes' incubation at 37°C sedimentation is achieved by  
15 centrifuging. The pellet, incubated for a further 2.5 hours at 37°C, is transferred into flat-bottomed wells. The syncytia (10-100 times larger than the initial cells) are counted with an overhead microscope. In the control samples, it is possible to see some 100-200  
20 syncytia in each well under the microscope.

The N-succinylated heparins that constitute the subject of the invention proved to be capable of inhibiting the formation of syncytia in concentrations of between 0.01 and 10 µg/well. Especially good results  
25 were obtained with the product as described in Examples 3 and 4. According to preliminary data, results equivalent to those obtained with the syncytia are obtained in cytofluorimetric conditions, where the N-succinylated heparins compete only with the bond of the  
30 anti-gp120 antibodies on the 8E51 cells. In view of their virtual lack of anticoagulant activity, the N-

WO 92/22588

PCT/EP92/01261

12

succinylated heparins according to the invention are proposed as selective antiviral agents that are substantially devoid of side effects.

5 For the envisaged therapeutic applications, the succinylated heparins according to the invention will be formulated in suitable pharmaceutical forms, using conventional methods and excipients, such as those described in Remington's Pharmaceutical Sciences Handbook, Mack Pub. Co., New York, U.S.A.. The heparins  
10 according to the invention will be administered for the most part parenterally, but the use of other administration routes, for example the oral route, is not ruled out, in which case appropriate salts would be used. The dosage will depend on various factors and  
15 will need to be established on an individual basis, but will generally be between 10 and 1000 mg of the compounds described above, given in one to four doses per day.

WO 92/22588

PCT/EP92/01261

13

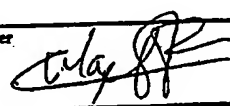
CLAIMS

1. N-acylated heparins with a level of acylation of between approximately 10% and approximately 100%.
- 5 2. Heparins according to claim 1, N-acylated with residues of aliphatic monocarboxylic acids with between 3 and 20 carbon atoms and aliphatic dicarboxylic acids with between 3 and 10 carbon atoms.
- 10 3. Heparins according to claim 2 in which the mono- and di-carboxylic acids can be substituted by one or more hydroxyl, amine or acylamine groups.
4. Use of a heparin with a level of N-succinylation of between 25% and 35% to prepare a drug effective in inhibiting the replication of retroviruses.
- 15 5. Process for the preparation of N-acylated heparins according to claims 1-3 from heparins, or their fractions, or their fragments, that are subjected to N-desulphation followed by N-acylation with a mono- or di-carboxylic acid or with their functional
- 20 derivatives.
6. Pharmaceutical compositions with anti-HIV activity containing as active principle an N-acylated heparin according to claims 1-3.

## INTERNATIONAL SEARCH REPORT

PCT/EP 92/01261

International Application No

<b>I. CLASSIFICATION OF SUBJECT MATTER</b> (If several classification symbols apply, indicate all) <sup>6</sup>		
According to International Patent Classification (IPC) or to both National Classification and IPC		
Int.Cl. 5 C08B37/10; A61K31/725		
<b>II. FIELDS SEARCHED</b>		
Minimum Documentation Searched <sup>7</sup>		
Classification System	Classification Symbols	
Int.Cl. 5	C08B ; A61K	
Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in the Fields Searched <sup>8</sup>		
<b>III. DOCUMENTS CONSIDERED TO BE RELEVANT<sup>9</sup></b>		
Category <sup>10</sup>	Citation of Document, <sup>11</sup> with indication, where appropriate, of the relevant passages <sup>12</sup>	Relevant to Claim No. <sup>13</sup>
X	US,A,3 118 816 (I. B. CUSHING) 21 January 1964 cited in the application see column 1, line 66 - column 2, line 2 see column 3, line 50 - column 4, line 26 ---	1,2,5
A	EP,A,0 293 826 (STICHTING REGA V. Z. W.) 7 December 1988 see page 5, line 48 - line 56; table 2 ---	1,5,6
A	WO,A,9 003 791 (DIFFERANTE) 19 April 1990 see page 3, line 25 - line 30 see page 5, line 30 - line 33; examples ---	1,6
<p><sup>10</sup> Special categories of cited documents:<sup>10</sup></p> <p>"A" document defining the general state of the art which is not considered to be of particular relevance</p> <p>"E" earlier document but published on or after the international filing date</p> <p>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</p> <p>"O" document referring to an oral disclosure, use, exhibition or other means</p> <p>"P" document published prior to the international filing date but later than the priority date claimed</p> <p>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</p> <p>"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step</p> <p>"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.</p> <p>"A" document member of the same patent family</p>		
<b>IV. CERTIFICATION</b>		
Date of the Actual Completion of the International Search	Date of Mailing of this International Search Report	
04 SEPTEMBER 1992	16. 09. 92	
International Searching Authority	Signature of Authorized Officer	
EUROPEAN PATENT OFFICE	MAZET J. 	

ANNEX TO THE INTERNATIONAL SEARCH REPORT  
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Patent document cited in search report	Publication date	Patent family member(s)	Publication date
US-A-3118816		None	
EP-A-0293826	07-12-88	JP-A- 1100127	18-04-89
WO-A-9003791	19-04-90	AU-A- 4426289	01-05-90

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